

THE SYNTHESIS OF THE RINGS A/B IN WITHA FERIN A AND OTHER WITHANOLIDES*

M. Weissenberg, E. Glotter and D. Lavie

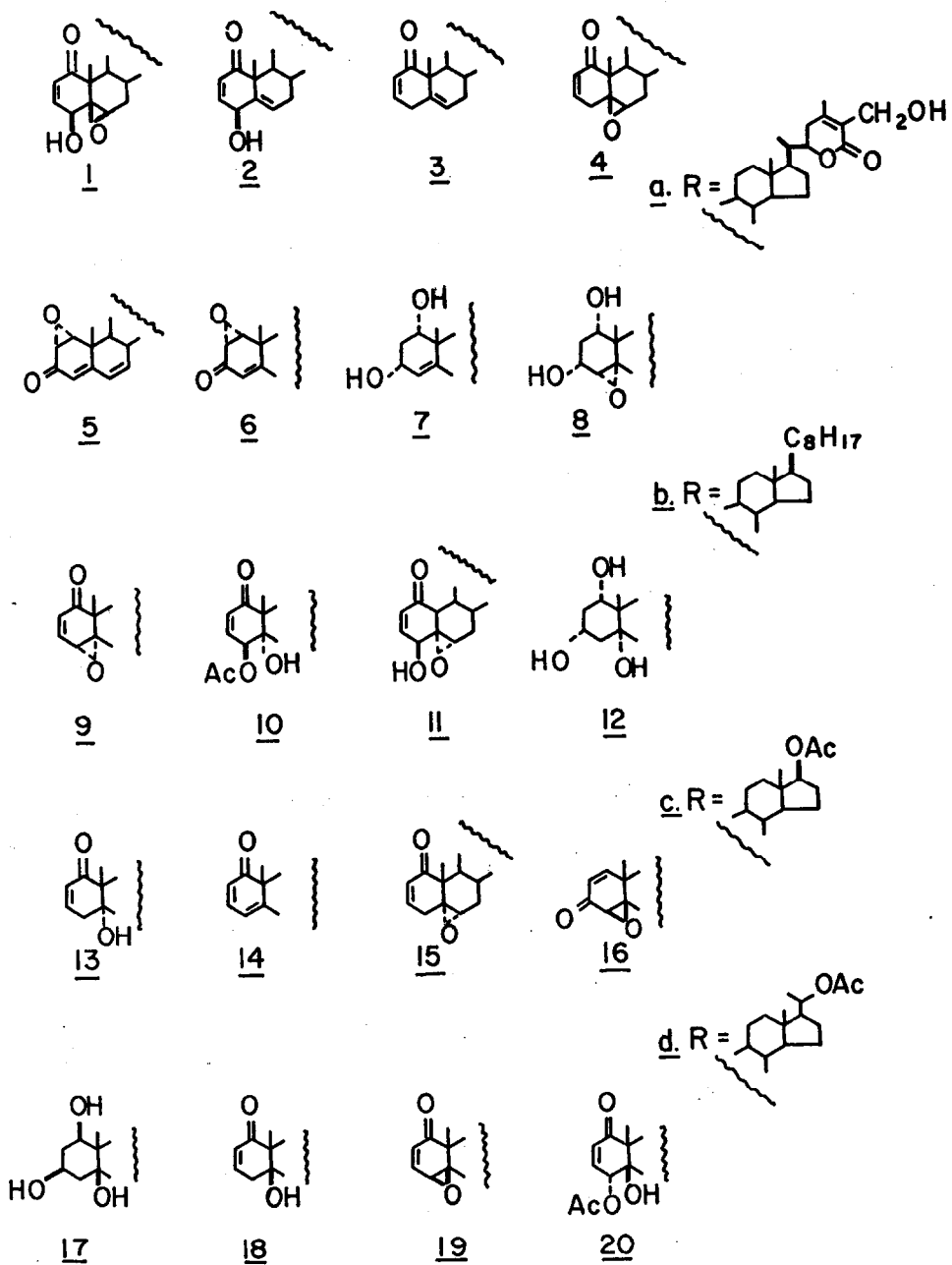
Department of Organic Chemistry, The Weizmann Institute of Science, Rehovot, Israel

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A recent communication on the synthesis of the A/B rings moiety of withaferin A¹, prompted us to report our own approach to this problem which is markedly different, and led to the successful preparation of the title derivatives. This study follows a detailed chemical investigation performed earlier in our laboratory² on the isolation and characterization of several withanolides^{2b}, a group of naturally occurring C₂₈ steroidal lactones identified in some species of Solanaceae³. Interest in the different physiological activities of withaferin A (1a)⁴ led us to consider synthetic routes for the construction of the characteristic A/B rings system found in withanolides. The present communication deals with the partial synthesis of four withanolide like substitution patterns (1-4) in the cholestane (1b-4b), 17 β -acetoxyandrostane (1c-4c) and 20 β -acetoxypregnane (1d-2d) series.

The sequence in the cholestane series starts with compound 5b obtained in two steps from cholesterol⁵. Catalytic hydrogenation (5% Pd-CaCO₃ in benzene) of 5b produced quantitatively the epoxy-enone 6b m.p. 118-120° which upon treatment with LiAlH₄ yielded the unsaturated diol 7b; peracid epoxidation gave stereoselectively the α -epoxide 8b. Selective acetylation at C-3 followed by mild oxidation at C-1 and elimination on alumina, yielded the epoxy-enone 9b (overall yield 70% from 5b). The trans junction of the A/B rings in 9b was proved by the positive solvent shift (Δ CDCl₃/C₆H₆ + 11.5 Hz) experienced in the nmr spectrum by the C-10 Me⁶. Opening of the epoxide ring of 9b in acidic conditions and subsequent acetylation led to compound 10b (75%), m.p. 125-126°, nmr Δ CDCl₃/C₆H₆ + 6 Hz. Dehydration of 10b (SOCl₂ in pyridine) followed by mild hydrolysis of the 4-acetate, gave quantitatively 2b, m.p. 125-127°, nmr δ (CDCl₃) 6.82 (3-H, dd, J=10 and 4.5 Hz), 6.05 (6-H, t), 5.99 (2-H, dd, J=10 and < 1 Hz), 4.64 (4-H, dd, J=4.5 and < 1 Hz), 1.45 (19-H, s)

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and 0.71 (18-H, s). The corresponding androstane and pregnane derivatives were prepared using a similar set of reactions; compounds 2b, 2c (m.p. 198-200°) and 2d (m.p. 85-87°) have the A/B rings structure appearing in the withanolide 2a³.

Stereoselective peracid epoxidation of 2b afforded quantitatively the β -epoxide 1b, m.p. 223-225° [α]_D + 40° (c, 0.63, CHCl₃), M⁺ 414, nmr δ (CDCl₃) 6.96 (3-H, q, J=10 and 6 Hz), 6.22 (2-H, d, J=10 Hz), 3.75 (4-H, d, J=6 Hz) 3.23 (6-H, d, J=2.5 Hz), 1.40 (19-H, s) and 0.67 (18-H, s); the large downfield solvent shift (Δ CDCl₃/C₆H₆ - 14 Hz) of the C-10 Me is in agreement with the boat conformation assigned to ring A in withaferin A by solvent shifts⁶ and X-ray⁷ analysis. Similarly 1c, m.p. 248-250°, and 1d have been prepared; they all have the A/B rings moiety of withaferin A^{2a}. When the peracid epoxidation was done on the acetate derivative of 2b, a 2:1 mixture of α - and β -epoxides was obtained from which the α -epoxide only could be isolated. Following hydrolysis 11b was obtained, m.p. 185-187°, [α]_D + 102° (c, 0.5, CHCl₃), nmr Δ CDCl₃/C₆H₆ + 5 Hz.

In another set of reactions, compound 6b was converted to the triol 12b by a procedure developed in the androstane series⁸ and including first reduction with NaBH₄, then stereoselective peracid epoxidation followed by treatment with LiAlH₄, in an overall yield of 75%. Selective acetylation at C-3 followed by mild oxidation at C-1 and subsequent elimination on alumina gave the enone 13b, m.p. 132-133°, nmr Δ CDCl₃/C₆H₆ + 12 Hz, Δ CDCl₃/C₅H₅N - 3 Hz (70% yield from 12b). Dehydration of 13b (SOCl₂ in pyridine) led to a 2:1 mixture of the dienone 14b⁹ and 3b, easily separated to give pure 3b, m.p. 102-103°, [α]_D - 26.5° (c, 0.8, CHCl₃), $\lambda_{\text{max}}^{\text{EtOH}}$ 222 nm (ϵ 8800), nmr δ (CDCl₃) 6.80 (3-H, dq, J=10, 4.5 and 2.5 Hz), 5.90 (2-H, dq, J=10, 2.5 and < 1 Hz), 5.61 (6-H, m), 1.23 (19-H, s) and 0.70 (18-H, s); 3c has been similarly prepared, m.p. 156-158°. Compounds 3b and c have the A/B rings structure encountered in withanolides F, G, H, J, L, and M^{2d}, and in physalins B and C¹⁰. Further peracid epoxidation of 3b led to a 2:1 mixture of the α - and β -epoxides 15b and 4b, which was separated to give 15b, m.p. 121-123°, nmr Δ CDCl₃/C₆H₆ + 17 Hz and 4b, m.p. 114-115°, M⁺ 398, nmr δ (CDCl₃) 6.82 (3-H, dq, J=10, 5 and 2 Hz), 6.03 (2-H, dd, J=10 and 3 Hz), 3.12 (6-H, d, J=2.1 Hz), 1.24 (19-H, s) and 0.67 (18-H, s), Δ CDCl₃/C₆H₆ - 4 Hz; in a similar way compounds 15c (m.p. 181-183°) and 4c (m.p. 192-193°) were prepared. The compounds 4b-c have the A/B rings moiety found in withanolide E^{2d} and jaborosalactone A¹¹. In both the cholestane and androstane series, the major product of this epoxidation was found to be the α -epoxide; the β -epoxide was predominant in our previous experiments done on the peracid epoxidation of withanolide L having the system 3, however, with an additional Δ ^{14,2d}.

The related compounds 17b-20b have been prepared through a similar sequence starting from 16b. The triol 17b was converted to the enone 18b (m.p. 194-196°, nmr Δ CDCl₃/C₅H₅N - 22 Hz) using the same sequence as described for 12 \rightarrow 13. Dehydration of 18b (SOCl₂ in pyridine) gave exclusively 14b. Peracid epoxidation of 14b afforded stereoselectively the β -epoxide 19b, m.p. 155-157°, nmr Δ CDCl₃/C₆H₆ - 5 Hz. The epoxide ring in 19b was opened as described

above for 9 \longrightarrow 10 to give 20b, m.p. 170-172°, nmr $\Delta\text{CDCl}_3/\text{C}_6\text{H}_6$ - 8 Hz. The preparation of these pairs of isomers by independent sequences constitutes as well additional proof for the configurational assignments based on the nmr data.

Further details on chemical and biological studies will be given in forthcoming publications.

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REFERENCES

1. M. Ishiguro, A. Kajikawa, T. Haruyama, M. Morisaki and N. Ikekawa, Tetrahedron Letters 1421 (1974).
2. (a) D. Lavie, E. Glotter and Y. Shvo, J.Chem.Soc. 7517 (1965);
(b) I. Kirson, E. Glotter, A. Abraham and D. Lavie, Tetrahedron **26**, 2209 (1970);
(c) I. Kirson, E. Glotter, D. Lavie and A. Abraham, J.Chem.Soc.(C) 2032 (1971);
(d) E. Glotter, I. Kirson, A. Abraham and D. Lavie, Tetrahedron **29**, 1353 (1973).
3. D. Lavie, Nobel Symposium **25** (G. Bendz and J. Santesson editors), Nobel Foundation Stockholm and Academic Press New York, 1973, p. 181.
4. S. Ben Efraim and A. Yarden, Antibiot.Chemoterapy **12**, 576 (1962); B. Shohat, S. Gitter and D. Lavie, Int.J.Cancer **5**, 244 (1970).
5. E. Glotter, M. Weissenberg and D. Lavie, Tetrahedron **26**, 3857 (1970).
6. E. Glotter and D. Lavie, J.Chem.Soc.(C) 2298 (1967).
7. A.T. McPhail and G.A. Sim, Ibid (B) 962 (1968).
8. G. Eggart, P. Keller, C. Lehmann and H. Wehrli, Helv.Chim.Acta **51**, 940 (1968).
9. H. Izawa, M. Morisaki and K. Tsuda, Chem.Pharm.Bull (Tokyo) **14**, 873 (1966).
10. T. Matsuura, M. Kawai, R. Nakashima and Y. Butsugan, J.Chem.Soc.(C) 664 (1970);
M. Kawai and T. Matsuura, Tetrahedron **26**, 1743 (1970).
11. R.Tschesche, M. Baumgarth and P. Welzel, Tetrahedron **24**, 5169 (1968).